

Cadmium toxicity in fish: An overview

Puneet Kumar^{1*} and Anu Singh²

¹Centre for Wildlife Conservation Management & Disease Surveillance,
Indian Veterinary Research Institute, Izatnagar- 243 122 (U.P.), India

²Laboratory of Immunocytochemistry, Bareilly College, Bareilly-243006 (U.P.), India

Abstract

Environmental pollutants have hazardous impact on living organism present on this planet. Environmental pollutants give bad impact on the health of livestock as well as human beings and have been categorized as heavy metals, pesticides etc. Cadmium and its compounds are most common environmental toxicants with potential for bioaccumulation and persistence in the body, and produce versatile biotic changes in the aquatic ecosystem. Monitoring toxic metal level in aquatic ecosystem (especially fish) is the need of the awareness from public health point of view. It concluded that our fishes also carry good quantity of cadmium. It is inferred from the study that selective removal of tissues that accumulate highest levels of cadmium *i.e.* kidney and liver might reduce the chances of toxicity due to this metal in human consumers and in animals fed with fish-meal.

Keywords: Cadmium toxicity, fish, heavy metals, immunotoxicity, nephrotoxicity

Introduction

Next to air, water is essential constituent of life support system and its quality play pivotal role in the maintenance of health. Being as an important natural resource it is used for many purposes especially for aquaculture, industry, irrigation and domestic needs. Abundant resource of water is available in our country. Unfortunately, rapid industrialization, fast growth in population and non-judicious use of natural resources has resulted into many fold increase in water pollution problem. Most of the 14 major rivers of India are victims of water pollution; Ganga and Yamuna ranking top among them (Ajmal *et al.*, 1985; Singh, 2001; Jain and Sharma, 2001; Kaushik *et al.*, 2003; Sharma, 2003 and Kumar *et al.*, 2007, 2008, 2009).

Industrial effluents are major source of water pollution besides sewage, agricultural discharges and other household residues. Industrial effluents contain a variety of toxic pollutants including suspended solids, organic compounds, inorganic compounds, pesticides and various toxic metal compounds. The chief source of contaminants are the industrial waste discharge, mining, agriculture, household waste disposal and fuel combustion (Woodling *et al.* 2001; Patra *et al.*, 2005 and Swarup *et al.*, 2006; Saxsena and Garg, 2011). Aquaculture is totally based on aquatic

ecosystem and accounts for substantial contribution to Indian economy and also provide livelihood to millions of people. Spectacular achievements have been made in the field of aquaculture by adapting modern techniques of fish culture and hence there is substantial growth in the production of culture fishes. Total fish production in India in year 2002-2003 was to the tune of 6.2 million metric tones (Dwivedi, *et al.* 2004). Fishes are highly nutritious and consumed as a delicacy food through out country (Prasad and Kumar, 2007). But addition of pollutants in water has impact on the aquatic ecosystem and poses adverse effect on the fish health. As a result fish production is generally encumbered and fish farmers face a great economic loss. Thus monitoring and understanding pathophysiology of various toxicants would be helpful in minimizing losses and providing safeguard to public health.

A variety of contaminants including toxic heavy metals (cadmium, copper, mercury and zinc) are reported to be ubiquitously present in rivers, reservoirs and are disadvantageous for aquatic organisms (Olsson, 1998). In general, they are not biodegraded and therefore, their bioaccumulation in fish, oyster, mussels, sediments and other components of aquatic ecosystems have been reported from all over the world. It appears that problem of heavy metals accumulation in aquatic organisms including fish needs continuous monitoring and surveillance owing to

*Corresponding author: puneetbiochem@gmail.com

biomagnifying potential of toxic metals in human food chain (Das and Kaviraj 2000; Laxi, 2005; Jayakumar and Paul 2006; Kumar *et al.*, 2007; Kumar *et al.*, 2008; Kumar *et al.*, 2009).

Physico-chemical property of cadmium

Pure cadmium is a soft, silver-white metal. The physical property of cadmium is atomic number-48, atomic weight-112.411, electro-negativity-1.5, crystal ionic radius (Principal valence state)-0.97, ionisation potential-8.993, oxidation state +2, electron configuration Kr 4d¹ 5S², density-8.64 g/cm³, melting point - 320.9°C and boiling point-765°C at 100 kPa. It is usually found as a mineral combined with other elements such as oxygen (cadmium oxide), chlorine (cadmium chloride), or sulfur (cadmium sulfate, cadmium sulfide).

Source of exposure

It is an element that occurs naturally in the earth's crust and got rank 7 of ASTDR's "Top 20 list" (ASTDR, 1999). Percentage of cadmium in the upper soil has been increasing because it is found in insecticides, fungicides, sludge, and commercial fertilizers which are routinely used in agriculture. Dental alloys, electroplating, motor oil, and exhaust are other sources of Cd pollution. Hence, anthropogenic activities have increased Cd magnification in the environment. 10% of total Cd in the environment is derived from natural sources, whereas remaining 90% is derived from anthropogenic activity (Okada *et al.*, 1997). Volcanic activity contributes about 62% of natural emissions and other natural sources include decaying of vegetation (25%) airborne soil particles (12%) and forest fire (2%).

It's non-corrosive and cumulative nature has made it very important due to its applications in electroplating or galvanizing. It is also used as colour pigment for paints, plastics, and as a cathode material for nickel-cadmium batteries. Anthropogenic activities like; smelting operations, use of phosphate fertilizers, pigment, cigarettes smokes, automobiles etc. have contributed to the entry of cadmium into human and animal food chain (WHO, 1992; Okada *et al.*, 1997; Kumar *et al.*, 2007). Presence of cadmium at higher concentration than the maximum allowable limits in water, vegetation and food have been reported by author (Agarwal and Raj, 1978; Khandekar *et al.*, 1980; Allen, 1995; Laxi, 2005; Kumar *et al.*, 2008; Asagba, 2010).

Higher level of Cd has also been detected in sewage sludge (rich in almost all nutrients and hence generally used as plant fertilizer), various vegetables (Roblenbeck *et al.*, 1999), animals feed and their tissues (Kumar *et al.*, 2007). Topsoil enrich in sludge contributes Cd accumulation in the blood, milk, hair, liver and kidney of sheep, goat, cow, buffalo (Brebner *et al.*, 1993; Swarup *et al.*, 2005; Balangatharathilagar *et al.*, 2006 and Patra *et al.*, 2007). In India, various levels of cadmium concentration have been reported to be present in aquatic ecosystem which is more

than 5ng/ml in the Yamuna river water at Agra, Delhi, Etawah and Mathura (Ajmal *et al.*, 1985) and 0.50-114.8 mg/kg in the Yamuna river sediments at Agra and Delhi but the water around the industrial areas have been found to contain higher levels of cadmium (Singh, 2001 and Kaushik *et al.*, 2003). Similarly, Hindon River (Uttar Pradesh) has also been contaminated with heavy metals including cadmium (Jain and Sharma, 2001 and Sharma, 2003). Moreover, high concentration of cadmium (70-100 ng/ml) has been detected in Bombay city (Agrawal and Raj, 1978), Lalbag pond water of Baroda city (Kannan, 1997) and edible tissues of fish and chicken in western UP market (Kumar *et al.*, 2006 and 2007; Burger, 2008).

Site for absorption in fish

In the fish, the possible areas of absorption of dissolved metals are the gills (respiratory tract), the intestine (ingestive intake) and the skin (transcutaneous uptake).

Molecular mechanisms of absorption

There are various mode of Cd uptake in aquatic organism, where it is most readily absorbed by organisms directly from the water in its free ionic form Cd (II) (AMAP 1998). Metal ions are usually absorbed through passive diffusion or carrier mediated transport over the gills while metals associated with organic materials are ingested and absorbed by endocytosis through intestine. It has been suggested that cadmium ions enter the chloride cells in the gills through calcium channels (Olsson, P.E., 1998). Once enter in the cells the metal is made available for the interaction with cytoplasmic components such as enzymes (causing toxic effects) and Metallothioneine (probably being detoxified). Although Metallothioneine is induced in the gills it does not appear to be as capable of sequestering the vast majority of accumulated Cd²⁺, as it is in the liver (Olsson and Hogstrand, 1987). The reason for this is believed to be due to the high affinity of Cd²⁺ for Ca²⁺ binding sites in the gills (Flick *et al.*, 1987), and it is also believed that Cd²⁺ binds to the active sites on the basolateral Ca²⁺-pump in chloride cells. It thus seems that Cd²⁺ enters the gills through Ca-channel on the apical side and is further translocated to the circulation interactions with Ca²⁺-ATPases on the basolateral side.

Interaction with other elements

Zinc increases the toxicity of cadmium to aquatic invertebrates. However, high calcium concentrations in water protect them from cadmium uptake by competing at uptake sites. It is very rare that only one toxic element, at a time, is released into the aquatic ecosystem. Most of the heavy metals interact with each other and also influenced by other ions (e.g. Ca²⁺, Mg²⁺, Na²⁺, Mn²⁺, Fe²⁺, Pb²⁺, S²⁺, Se²⁺ and Ni²⁺). Calcium has been shown to interact with Cd²⁺ to potentiate or minimize their toxicity. Elevated ingestion Cd²⁺

can produce deficiency states of both Cu^{2+} and Zn^{2+} . Exposure of animals to Cd^{2+} results in alteration in the Zn^{2+} metabolism.

Tissue distribution

Bioaccumulation of cadmium takes place at tropic level and found to be highest in algae (Ferard *et al.*, 1983; Pinto *et al.*, 2003). It also accumulates in considerable concentrations in various organs of fish (Sindayigaya *et al.*, 1994; Kumar *et al.*, 2006; Kumar *et al.*, 2008). Smet & Blust (2001) reported that cadmium accumulates in tissues of carp *Cyprinus carpio* in following order: kidney > Liver > Gills. Kumar *et al.* (2005) have also reported similar accumulation pattern in *Clarias batrachus* in an experimental study. Some insects can also accumulate high levels of cadmium without showing any adverse effects (Jamil and Hussain, 1992). Kidney is the prime target organ for cadmium. The liver also stores a considerable part of the accumulated cadmium. Cadmium is redistributed to these organs directly following uptake through the gills and intestine, but there may also be redistribution of cadmium from other organs (Olsson and Hogstrand, 1987).

Toxic effect

Cadmium has been reported to exert deleterious effects in terms of nephrotoxic, cytotoxic, genotoxic, immunotoxic and carcinogenic (ASTDR, 1999; Lippmann, 2000 and Rissode-faverney, 2001).

1. Nephrotoxicity

Cadmium is heavy metal and poses high toxicity at very low level of exposure and has acute and chronic effects on aquatic animal health and environment. Long exposure of cadmium produces a wide variety of acute and chronic effects in aquatic animals. It's prime site is kidney (Thomas *et al.*, 1983 and Kuroshima, 1992). According to the current knowledge kidney damage (renal tubular damage) is probably the critical health effect (Jarup *et al.*, 1998). Not only this it also creates disturbances of calcium metabolism, hypercalciuria and takes part in the formation of stones in the kidney. The toxicity is variable in fish, salmonoids being particularly susceptible to cadmium. Sublethal effects in fish, notably malformation of the spine, have been reported.

2. Induction of Oxidative Stress

Free radicals and other reactive oxygen species (ROS) have been recently incriminated in the pathogenesis of various metal toxicities (Yiin *et al.*, 1999a and 1999b; Senapati *et al.*, 2001; Basha and Rani, 2003; Rahman, 2003; Suresh, 2009). There are many reports suggesting alterations in free radicals production and antioxidant defense system of the body after cadmium exposure. Administration of Cadmium chloride at the dose rate of $15\mu\text{g/ml}$ in drinking water for 30 days revealed significant increase in lipid peroxidation (LPO) in cortical region of kidney (Oner *et al.*, 1995). Treatment of rats with Cd^{2+} significantly increases in LPO in heart within

3 hours of Cd^{2+} injection and kidney and liver within 6-12 hours. Superoxide dismutase (SOD) activity increased in heart, liver and kidney within 24 hours of Cd^{2+} intoxication. Catalase activities were also increased significantly in heart after 9 hours of Cd^{2+} injection without any significant change in liver and kidney (Sarkar *et al.*, 1995).

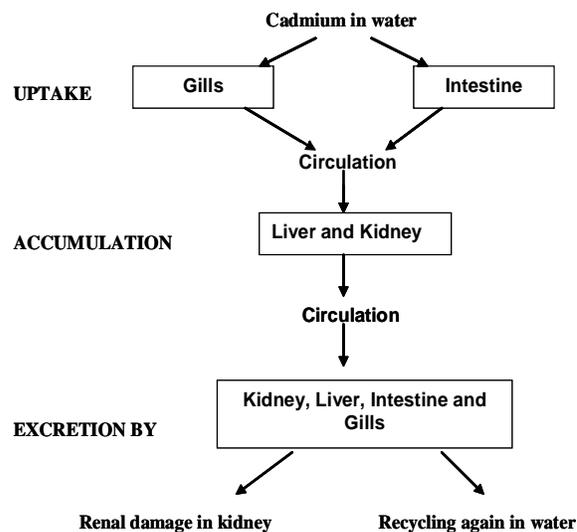


Fig. : 1 The cycle of Cadmium in aquatic ecosystem

3. Immunotoxicity

A number of investigations have suggested that cadmium may exert immunosuppressive effects of cadmium exposure in both fishes and mammals (Zellkoff *et al.*, 1995; Kim *et al.*, 2000 and Giari *et al.*, 2007). Recent reports suggest that cell mediated immunity is most affected (Kumar *et al.*, 2008) and phagocytosis, natural killer cell activity and host resistance towards experimental infections are markedly impaired in cadmium toxicity. Sovenyl and Szkolczal (1993) also reported marked immunosuppressive effects of cadmium exposure on common carp in terms of lowered antibody response, lysozyme level and microcidal capacity of phagocytes. Some reports suggest that cadmium enhances humoral immune response at low level of exposure (Descotes, 1992; Krumschnabel *et al.*, 2010).

4. Effect on organ structure and function

Cadmium in high doses induce structural and function alterations in various vital organs including liver, kidney, gill and intestine of fishes.

Liver: Cadmium accumulates in liver of fishes in high concentrations (Smet and Blust, 2001; Rangsayatorn *et al.*, 2004). It also induces various pathological changes in liver tissues including engorgement of blood vessels, congestion, vacuolar degeneration of hepatocytes, necrosis of pancreatic cells and fatty changes in the peripancreatic hepatocytes (Rani and Ramamurthi, 1989; Dangre *et al.*, 2010).

Kidney: Cadmium accumulates in kidney of fishes in maximum concentration. Cadmium has been reported to possess nephrotoxic action in man and various animals. In fact, kidney is the principle target organ of cadmium toxicity and chronic cadmium exposure in almost all animal species is characterized by varying degree of renal damage (Roméo *et al.*, 2000; Shukla and Gautam, 2004; Kumar *et al.*, 2006; Kumar *et al.*, 2009; Vesey, 2010).

Gills: Gills are also reported to act as storehouse of cadmium in experimental studies (Allen, 1995; Tao *et al.*, 2000; Fafioye *et al.*, 2004; Ramesh and Nagarajan, 2007). Wong and Wong (2000) studied morphological and biochemical changes in the gills of Tilapia (*Oreochromis mossambicus*) after experimental cadmium exposure. In scanning electron microscopic studies, they found an augmentation of microbridges in pavement cells and an increase in the apical membrane of chloride cells. They further reported chloride cells as a prime target of cadmium toxicity, resulting into fish hypocalcemia.

Other organs like intestine and gonads of fishes also appear susceptible for ill effects of cadmium toxicity (Taylor, 1983; Kumari and Ram, 1997; Singh *et al.*, 2007; Kumar, 2007).

Conclusion

Cadmium enters into the aquatic ecosystem through anthropogenic activity and gets further biomagnified in the food chain. Studies revealed the high concentration in of Cadmium in kidney in comparison to the liver and disrupts the normal calcium metabolism resulting into hypocalcaemia and hyperglycemia. It is vivid that it can be concluded that fishes are the major concern to increase the cadmium in their haematopotic organs *viz.* kidney and liver. This study indicates that high levels of Cd in fish are potential risk concern on human consumer's health.

References

1. Agrawal VK and Raj KPS (1978). Metal contents in the drinking water of Cambay. *Air and Soil Pollution*. 9: 129-134.
2. Ajmal M, Khan MA and Nomani AA (1985). Distribution of heavy metals in plants and fish of the Yamuna river. *Environ. Monitoring Assess.* 5: 361.
3. Allen P (1995). Chronic accumulation of cadmium in the edible tissues of *Oreochromis aureus* (Steindachner): modification by mercury and lead. *Arch. Environ. Con. Toxicol.* 29(1): 8-14.
4. AMAP (1998). Assessment report: Arctic pollution issues. Arctic Monitoring and Assessment Programme, Oslo.
5. Asagba SO (2010). Comparative effect of water and food-chain mediated cadmium exposure in rats. *Biometals*. (Article in press)
6. ATSDR (1999). Toxicol. Profile of Cadmium. Agency for Toxic Substances and Drug Registrar, Atlanta, GA. US Deptt. of Health and Human Services.
7. Balagangatharathilagar M, Swarup D, Patra RC and Dwivedi SK (2006). Blood lead level in dogs from urban and rural areas of India and its relation to animal and environmental variables. *Sci. Total Environ.* 359:130-134
8. Basha PS and Rani AU (2003). Cadmium induced antioxidant defense mechanism in freshwater teleost *Oreochromis mossambicus* (Tilapia). *Ecotoxicol. and Environ. Safety.* 56(2): 218-221.
9. Burger J (2008). Assessment and management of risk to wildlife from cadmium. *Sci. Total Environ.* 389: 37 – 45.
10. Dangre AJ, Manning S and Brouwer (2010). Effects of cadmium on hypoxia-induced expression of hemoglobin and erythropoietin in larval sheepshead minnow, *Cyprinodon variegates*. *Aquatic Toxicol.* 99(2):168-175.
11. Das S and Kaviraj A (2000). Cadmium accumulation in different tissues of common carp, *Cyprinus carpio* treated with activated charcoal, EDTA and single superphosphate. *Geobios.* 27:69-72.
12. Descotes J (1992). Immunotoxicology of cadmium. *IARC Science Publication.* 118: 385-390.
13. Dwivedi SN, Bhaumik U, Paria T, Saha SK and Mitra A (2004). Aided invasion of Exotic catfishes towards Indian Aquaculture. *Fishing Chimes.* 24(1): 106-109.
14. Fafioye OO, Adebisi AA and Fagade SO (2004). Toxicity of *Parkia biglobosa* and *Raphia vinifera* extracts on *Clarias gariepinus* juveniles. *African J. Biotechnol.* 3(11): 627-630.
15. Ferard JE, Jouany JM, Truhant R and Vasseur P (1983). Accumulation of cadmium in freshwater food chain experimental model. *Ecotoxicol. and Environ. safty.* 7(1): 43-52.
16. Flick G, Van Rijis JH and Wendelaar Bonga SE (1987). Evidence for high-affinity Ca⁺²-ATPase activity and

- ATP driven Ca^{+2} -transport in membrane preparations of the gill epithelium of the cichlid fish *Oreochromis mossambicus*. *J. Experi. Biol.* 119: 335-347.
17. Giari L, Manera M, Simoni E and Dezfuli BS (2007). Cellular alterations in different organs of European sea bass *Dicentrarchus labrax* (L.) exposed to cadmium. *Chemosphere*. 67(6): 1171-1181.
 18. Jain CK and Sharma MK (2001). Distribution of trace metals in the Hindon river system, India. *J. Hydrology Amsterdam*. 253(1-4): 81-90.
 19. Jamil K and Hussain S (1992). Biotransfer of metals to the insect *Neochetina eichhornae* via aquatic plants. *Archives Environ. contamin. and Toxicol.* 22: 459-463.
 20. Järup L, Berglund M, Linder G, Nordberg G and Vahter M (1998). Health effects of cadmium exposure - a review of the literature and a risk estimate. *Scandain J. Work Environ. Health*. 24 (1):52p.
 21. Jayakumar P and Paul VI (2006). Patterns of cadmium accumulation of the catfish *Clarias batrachus* (Linn.) exposed to sublethal concentration of cadmium chloride. *Veterinarshki Archiv*. 76:167-177.
 22. Kannan K (1997). Metals and metallic compounds In: *Fundamental of Environmental Pollution*. S. Chand & Company Publication. N. Delhi, India. pp: 142-183.
 23. Kaushik A, Jain S, Dawra J and Sharma P (2003). Heavy metal pollution in various canals originating from river Yamuna in Haryana. *J. Environ. Biol.* 24(3): 331-337.
 24. Khandekar RN, Kelkar DN and Vohra KG (1980). Lead, cadmium, copper, zinc and iron in atmosphere of greater Bombay. *Atmosphere Res.* 14: 457-461.
 25. Kim YO, Ahn YK and Kim JH (2000). Influence of melatonin on immunotoxicity of cadmium. *Int. J. Immunopharmacol.* 22(4): 275-284.
 26. Krumschnabel G, Ebner HL, Hess MW and Villunger A (2010). Poptosis and necroptosis are induced in rainbow trout cell lines exposed to cadmium. *Aquatic Toxicol.* 99(1):73-85.
 27. Kumar P, Prasad Y, Dhama K and Nandi D (2007). *In vitro* lymphoproliferative responses of peripheral blood mononuclear cells from different *in vivo* treated catfish (*Claris batrachus*). *Indian J. Comparative Microbiol., Immunol. Infectious Disease*. 27(1&2): 27-31.
 28. Kumar P, Prasad Y, Patra AK and Swarup D (2007). Levels of Cadmium and Lead in Tissues of Freshwater Fish (*Clarias batrachus* L.) and Chicken in Western UP (India). *Bull. Environ. Contamin. and Toxicol.* 79: 396-400.
 29. Kumar P, Prasad Y, Patra AK, Ranjan R, Patra RC, Swarup D and Singh SP (2009). Ascorbic acid, garlic extract and taurine alleviate cadmium-induced oxidative stress in freshwater catfish (*Clarias batrachus*). *The Sci. Total Environ.* 407: 5024–5030.
 30. Kumar P, Prasad Y, Ranjan R, Swarup D, Pattanaik AK and Patra RC (2008). Accumulation Pattern of Cadmium in Tissues of Indian Catfish *Clarias batrachus*. *Animal Nutrition. and Feed Technol.* 8(1): 115-119.
 31. Kumar P, Prasad Y, Sharma R and Patil RD (2006). Histopathological changes in liver and kidney of *Clarias batrachus* due to cadmium toxicity and its neutralization by Vitamin – C, Taurine and Garlic. *Proceedings of an national conference held in Chennai, 27 – 29 Dec 2006*. pp-42.
 32. Kumar P, Prasad Y, Swarup D, Patra RC and Nandi D (2005) Bioaccumulation of cadmium in fresh water indian catfish *Clarias batrachus*. *Proceedings of an international conference held in Lucknow (India), 14-17th November 2005*. pp: 45.
 33. Kumar P, Prasad Y, Raikwar MK, Singh M and Sharma AK (2006). Cadmium residues in the fish offals of Bareilly market region. *Proceedings of an international conference held in Palampur, 12 – 14 October 2006*. pp-114.
 34. Kumari SA and Ramkumar NS (1997). Effect of water pollution on histology of intestine of two fresh water fishes from Hussainsagar lake (A.P.). *Indian J. Environ. and Toxicol.* 7(2): 68-70.
 35. Kuroshima R (1992). Cadmium accumulation in the mummichog, *Fundulus heteroclitus*, adapted to various salinities. *Bull. Environ. Contam. Toxicol.* 49: 680-685.
 36. Laxi R (2005). Cadmium contamination in common Indian food items, *Himalayan J. Environ. Zool.* 19-23.

37. Lippmann M (2000). Human exposures and their health effects. In: *Environmental Toxicants* (2nd Edition) Wiley Intersciences. USA. pp-824-829.
38. Okada IA, Sakuma AM, Maio FD, Dovidemskas S and Zenebon O (1997). Evaluation of lead and cadmium in milk due to environmental contamination in Paraiba Valley region of South Eastern Brazil. *Revista-de-Saude-Publication*. 31:140-143.
39. Olsson PE and Hogstrand C (1987). Subcellular distribution and binding of cadmium to metallothionein in tissues of rainbow trout after exposure to 109Cd via the water. *Environ. Toxicol. Chem.* 6: 867-874.
40. Olsson PE (1998). Disorders associated with heavy metal pollution. In: *Fish Diseases and Disorders Volume 2 (Non-infectious Disorders)*. (Eds. Leatherland, J.E. and Woo, P.T.K.), CABI International, U.K. pp. 105-131.
41. Oner G, Senturk UK and Izgut UN (1995). The role of cadmium in the peroxidative response of kidney to stress. *Biol. Trace Elem. Res.* 48: 111-117.
42. Patra RC, Swarup D, Naresh R, Puneet K and Shekhar P (2005). Cadmium level in blood and milk from animals reared around different polluting sources in India. *Bull. Environ. Contam. Toxicol.* 76(4): 1092-1097.
43. Patra RC, Swarup D, Naresh R, Kumar P, Nandi D, Shekhar P, Roy S and Ali SL (2007). Tail hair as an indicator of environmental exposure of animals to lead and cadmium in different industrial areas. *Ecotoxicol. Environ. Saf.* 66(1): 127-131.
44. Pinto E, Sigaud Kutner TCS, Leitao MAA, Okamoto OK, Morse D and Colepicolo P (2003). Heavy metal-induced oxidative stress in algae. *J. Phycology*. 39(6): 1008-1018.
45. Prasad Y, Kumar P and D Nandi (2007). Healthful Perspectives of Fish. *Vererinary World* (Article in press).
46. Rahman K (2003). Garlic and aging: new insights into an old remedy. *Ageing Res. Rev.* 2: 39-56.
47. Ramesh F and Nagaranjan K (2007). Histopathological changes in gills of *Clarias batrachus* treated with sago effluent. *J. Exp. Zool.* 10(1): 169-171.
48. Rangsayatorn N, Kruatrachue M, Pokethitiyook P, Upatham ES, Lanza GR and Singhakaew S (2004). Ultrastructural changes in various organs of the fish *Puntius gonionotus* fed cadmium-enriched cyanobacteria. *Environ. Toxicol.* 19(6):585-93.
49. Rani UA and Ramamurthi R (1989). Histopathological alteration in the liver of freshwater teleost *Tilapia mossambica* in response to cadmium toxicity. *Ecotoxicol. Environ. Saf.* 17(2): 221-216.
50. Risso-de-faverney C, Devaus A, Lafaurie M, Girard JP, Bailly B and Rahmani R (2001). Cadmium induces apoptosis and genotoxicity in rainbow trout hepatocytes through generation of reactive oxygen species. *Aquatic Toxicol. Amsterdam.* 53(1): 65-76.
51. Roblenbeck SI, Burberg A and Zimmermann RD (1999). Lead and cadmium in Ethiopian vegetables. *Bull. Environ. Contam. Toxicol.* 62: 30-33.
52. Romeo M, Bennani N, Gnassia-Barelli M, Lafaurie M and Girard JP (2000). Cadmium and copper display different responses towards oxidative stress in the kidney of the sea bass *Dicentrarchus labrax*. *Aquatic Toxicol.* 48: 185-194
53. Sarkar S, Yadav P, Trivedi R, Bansal AK and Bhatnagar D (1995). Cadmium induced lipid peroxidation and the status of the antioxidant system in rat tissues. *J. Tarce Elem. Med. Biol.* 9: 144-149.
54. Saxsena R and Garg P (2011). Vitamin E provides protection against *In vitro* oxidative stress due to pesticide (Chlorphrifos and Endosulfan) in goat RBC. *GERF Bull. Biosci.* (Article in press).
55. Senapati SK, Dey S, Dwivedi SK and Swarup D (2001) Effect of garlic (*Allium sativum* L.) extract on tissue lead level in rats. *J. Ethnopharmacol.* 76: 229-232.
56. Sharma PD (2003). Environmental Pollution. In: *Ecology and Environment* (7th Edition) Rastogi Publication. Meerut, India. pp: 415-489.
57. Shukla S and Gautam RK (2004). Histopathological changes in the kidney of *Clarias batrachus* exposed to nuvan. *Flora and Fauna.* 10(1): 39-40.
58. Sindayigaya E, Canwnbergh RV, Robberecht H and Deelstra H (1994). Copper, zinc, manganese, iron, lead, cadmium, mercury and arsenic in fish from lake Tanganyika, Burundi. *Sci. Total Environ.* 144: 103-115.

59. Singh AP, Singh AK and Singh JPN (2007). Cadmium induced changes on the secretion of branchial mucous cells of peppered loach, *Lepdocephalichthys guntea*. *J. Exp. Zool.* 10(1): 65-68.
60. Singh M (2001). Heavy metal pollution in freshly deposited sediments of the Yamuna river (the Ganges River tributary): A case study from Delhi and Agra Urban centres, India. *Environ. Geol. (Berlin)*. 40(6): 664-671.
61. Sovenyl J and Szakolczal J (1993). Studies on the toxic and immunosuppressive effects of cadmium on the common carp. *Acta Vet. Hung.* 41(3-4): 415-426.
62. Sumet HDe and Blust R (2001). Stress responses and changes in protein metabolism in carp *Cyprinus carpio* during cadmium exposure. *Ecotoxicol. Environm. Saf.* 48(30): 255-262.
63. Suresh N (2009). Effect of cadmium chloride on liver, spleen and kidney melano macrophage centres in *Tilapia mossambica*. *J. Environ. Biol.* 30(4):505-8.
64. Swarup D, Patra RC, Naresh R, Kumar P and Shekhar P (2005). Blood lead levels in lactating cows reared around polluted localities; transfer of lead in to milk. *Sci. Total Environ.* 347: 106-110.
65. Swarup, D, Patra RC, Naresh, Ram, Puneet K., Pallav S, and Balagangatharathilagar M. (2006). Deficiency of copper and cobalt in goats reared around lead-zinc smelter. *Small Ruminant Res.* 63(3): 309-313.
66. Tao S, Liu C, Dawson R, Long A and Xu F (2000). Uptake of cadmium adsorbed on particulates by gills of goldfish. *Ecotoxicol. and Environ. Safety.* 47(3): 306-313.
67. Taylor D (1983). The significance of the accumulation of cadmium by aquatic organisms. *Ecotoxicol. and Environ. Safety.* 7(1): 33-42.
68. Thomas DG, Solbe JF, de LG, Kay J and Cryer A (1983). Environmental cadmium is not sequestered by metallothionein in rainbow trout. *Biochem. and Biophysic. Res. Communi.* 110: 584-592.
69. Vesey DA (2010). Transport pathways for cadmium in the intestine and kidney proximal tubule: Focus on the interaction with essential metals. *Toxicol. Letters.* 198(1):13-19.
70. WHO (1992). Environmental Health Criteria, No. 134, Environmental aspects, Geneva WHO.
71. Wong CK and Wong MH (2000). Morphological and biochemical changes in the gills of *Tilapia (Oreochromis mossambicus)* to ambient cadmium exposure. *Aquatic Toxicol.* 48(4): 517-527.
72. Woodling JD, Brinkman SF, Horn BJ (2001). Non uniform accumulation of cadmium and copper in kidney's of wild brown trout *Salmo trutta* populations. *Arch. Environ. Contam. Toxicol.* 40:381-385
73. Yiin SL, Chern CL, Sheu JY and Lin TH (1999a). Cadmium induced lipid peroxidation in rat tests and protection by selenium. *Biometals.* 12:365-359.
74. Yiin SL, Chern CL, Sheu JY, Tseng WC and Lin TH (1999b). Cadmium induced renal lipid peroxidation in rat and protection by selenium. *J. Toxicol. Environ. Health.* 57:403-413.
75. Zellkof JT, Brwser D, Squibb KS and Frenkel K (1995). Immunotoxicity of low level cadmium exposure in fish: an alternative animal model for immunotoxicological studies. *J. Toxicol. Environ. Health.* 45(3): 235-248.